KIF18A Motor Domain (1-374) His-Protein: wild-type (Human recombinant)
Cat. # CS-KF18  
Lot # 015  
1 x 100 µg  
Upon arrival store at 4°C (desiccated)  
See datasheet for storage after reconstitution

**Material**
The wild-type human motor domain of the kinesin-like KIF18A protein has been produced in a bacterial expression system. The recombinant protein contains a 6x His-tag at the amino terminal end and amino acids 1-374. The molecular weight of KIF18A is approximately 44 kDa and it is supplied as a white lyophilized powder.

**Storage and Reconstitution**
Before reconstitution, briefly centrifuge to collect the product at the bottom of the tube. The protein should be reconstituted to 5 mg/ml with the addition of 20 µl of nanopure water (100 µg size). When reconstituted, the protein will be in the following buffer: 100 mM Tris pH 8.0, 100 mM NaCl, 2 mM MgCl₂, 15% (w/v) sucrose, and 1% (w/v) dextran. In order to maintain high biological activity of the protein, it is strongly recommended that the protein solution be supplemented with DTT to 1 mM final concentration, aliquoted into "experiment-sized" amounts, snap frozen in liquid nitrogen, and stored at -70°C. The protein is stable for six months if stored at -70°C. Defrosting should be in a room temperature water bath for 3 min then place in ice. Dilution of protein must be done in 100 mM Tris pH 8.0 and 100 mM NaCl buffer. The protein should not be exposed to repeated freeze-thaw cycles. The lyophilized protein is stable at 4°C desiccated (<10% humidity) for one year.

**Purity**
Protein purity is determined by scanning densitometry of Coomasie Blue-stained protein on a 4-20% polyacrylamide gradient gel. KIF18A 1-374 protein was determined to be 80% pure. (see Figure 1).

**Biological Activity**

**Microtubule activated ATPase Assay**
KIF18A ATPase activity was measured by monitoring real time free phosphate generation using the Kinesin ELIPA Assay Kit (Cat. # BK060). The assay is based upon an absorbance shift (330 nm to 360 nm) that occurs when 2-amino-6-mercapto-7-methylpurine ribonucleoside (MESG) is cataclytically converted to 2-amino-6-mercapto-7-methylpurine in the presence of inorganic phosphate (Pi). One molecule of Pi will yield one molecule of 2-amino-6-mercapto-7-methylpurine in an essentially irreversible reaction. Hence, the absorbance at 360 nm is directly proportional to the amount of Pi generated in the kinesin ATPase reaction. Under the conditions outlined below, the Vmax for KIF18A microtubule-activated ATPase activity is >1500 nmoles ATP generated per minute per mg of KIF18A (Figure 2).

**Reagents**
1. Kinesin ELIPA Assay Kit (Cat.# BK060)
2. Recombinant KIF18A protein (Cat.# CS-KF18)

**Equipment**
Monochromatic spectrophotometer (set to 360 nm) or a filter based spectrophotometer with a 360 nm filter and bandwidth of <10 nm.

**Method (ELIPA ATPase assay)**
1. Thaw out and aliquot ELIPA reaction buffer, ELIPA 1 and 2 reagents, microtubules, taxol, and ATP reagents as described in the BK060 datasheet (https://www.cytoskeleton.com/bk060).
2. Resuspend 1-vial of KIF18A to 1 mg/ml with 100 µl 100 mM Tris-HCl pH=8.0 and 100 mM NaCl buffer.
3. Mix the components together IN THE ORDER SHOWN IN TABLE 1 BELOW.

<table>
<thead>
<tr>
<th>Order</th>
<th>Reagent</th>
<th>Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>ELIPA Reaction Buffer (room</td>
<td>2 ml</td>
</tr>
<tr>
<td></td>
<td>temperature)</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Taxol stock</td>
<td>20 µl</td>
</tr>
<tr>
<td>3</td>
<td>MT’s</td>
<td>160 µl</td>
</tr>
<tr>
<td>4</td>
<td>ELIPA Reagent 1</td>
<td>480 µl</td>
</tr>
<tr>
<td>5</td>
<td>ELIPA Reagent 2</td>
<td>24 µl</td>
</tr>
</tbody>
</table>

4. Gently swirl MT ELIPA MIX to mix evenly and incubate for 5 minutes at room temperature.
5. The MT ELIPA MIX is now ready for the addition of your motor protein.

**Figure 1. KIF18A 1-374 Protein Purity Determination.** A 10 µg sample of recombinant KIF18A protein (molecular weight approx. 44 kDa) was separated by electrophoresis in a 4-20% SDS-PAGE system and stained with Coomasie Blue. Protein quantitation was determined using the Precision Red Protein Assay Reagent (Cat. # ADV02). Mark12 molecular weight markers are from Invitrogen.
6. Aliquot 1 ml of MT ELIPA Mix to a 1.5 ml Eppendorf tube. This will be the no motor control.
7. Aliquot 1.5 ml of MT ELIPA Mix to 1.5 ml Eppendorf tube. Add 1 µl 1 mg/ml KIF18A to 1.5 ml MT ELIPA Mix and mix.
8. Aliquot 300 µl of no motor control to well positions A1-B1 in a 96-well plate.
9. Aliquot 300 µl of KIF18A MT ELIPA mix to well positions C1-D1 in a 96-well plate.
10. Setup the spectrophotometer in kinetic mode to take readings every 30 seconds for 30 min at room temperature.
11. Insert plate into spectrophotometer and start kinetic run.
12. After 3 mins or when baseline stabilizes, interrupt run, and aliquot 27 µl of working stock 10 mM ATP into each well with a multichannel pipettor to begin reactions simultaneously.
13. Immediately read the reactions again at 360 nm.

Figure 2. KIF18A microtubule ATPase activity determined with Kinesin ELIPA Kit (Cat#. BK060). Figure legend: KIF18A (0.2 µg) ATPase activity was measured (n=5) at an absorbance of 360 nm on an iD5 multi-mode microplate reader ( Molecular devices) over 30 minutes at room temperature. The ATPase activity was started when 0.9 mM of ATP was added 3 minutes into the kinetic run. Control reactions (n=5) were carried out in the absence of KIF18A motor protein.

Microtubule Activated Endpoint ATPase Assay
KIF18A ATPase activity was determined by measuring the inorganic phosphate (Pi) generated during the microtubule activated ATPase activity with the HTS Kinesin ATPase Endpoint Assay Biochem Kit (Cat.# BK053). This is a fast and economical method suitable for HTS applications. The assay is based on up a colorimetric change, measure at 650 nm.

Reagents
1. HTS Kinesin ATPase Endpoint Assay Biochem Kit (Cat.# BK053)
2. Recombinant KIF18A protein (Cat.# CS-KF18)

Method
1. Thaw out and aliquot kinesin reaction buffer, microtubules, taxol, kinesin motor protein, and ATP reagents as described in the BK053 datasheet (https://www.cytoskeleton.com/bk053).
2. Resuspend 1-vial of KIF18A to 0.5 mg/ml with 100 µl 100 mM Tris-HCl pH=8.0 and 100 mM NaCl buffer.
3. Transfer 240 µl 0.5 mg/ml KIF18A 1-374 to a new 1.5 ml centrifuge tube with 510 µl 100 mM Tris-HCl pH=8.0 and 100 mM NaCl buffer to produce a 0.16 mg/ml KIF18A working stock.
4. Aliquot reagents as described in Table 2.

Table 2. MT Activated ATPase Reactions.

<table>
<thead>
<tr>
<th>Well Position</th>
<th>Kinesin Reaction Buffer + Taxol</th>
<th>MTs (0.2mg/ ml)</th>
<th>KIF18A (0.16 mg/ml)</th>
<th>Well Designation</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1</td>
<td>30 µl</td>
<td>0</td>
<td>0</td>
<td>Blank</td>
</tr>
<tr>
<td>B1</td>
<td>20 µl</td>
<td>10 µl</td>
<td>0</td>
<td>MT only</td>
</tr>
<tr>
<td>C1</td>
<td>28.75 µl</td>
<td>0</td>
<td>1.25 µl</td>
<td>KIF18A only</td>
</tr>
<tr>
<td>D1</td>
<td>18.75 µl</td>
<td>10 µl</td>
<td>1.25 µl</td>
<td>KIF18A ATPase</td>
</tr>
</tbody>
</table>

5. Before adding the ATP, make sure spectrophotometer is set up correctly in end-point read mode at 650 nm.
6. To start the reactions, use a 8-well multichannel pipettor to aliquot 5 µl of ATP per reaction.
7. Allow the reactions to proceed at room temperature for 10 minutes.
8. Terminate reaction by adding 70 µl of CytoPhos to each well.
9. Allow the stop reaction to proceed for 10 minutes and take readings at 650 nm. Use well A1 (Table 2) as your blank.

Figure 3. KIF18A ATPase activity determined with HTS Kinesis ATPASE Endpoint Assay Biochem Kit (Cat#. BK053). Figure legend: KIF18A (0.2 µg) ATPase activity was measured on a spectrophotometer at an absorbance of 650 nm in a 96-well half-area plate at room temperature.

Product Uses
- Measurement of microtubule-activated ATPase assays
- Identification/characterization of proteins or small molecules that affect motor ATPase activity.
- Identification/characterization of proteins or small molecules that affect motor/microtubule interactions.

References

Product Citations/Related Products
For the latest citations and related products please visit www.cytoskeleton.com.