



Helping advance science,
one protein at a time.

How the RhoA/ROCK Pathway Changes with Age and its Connection to the Diseases of Aging

Related Publications Research Tools

December 2024

Sponsored Conferences

ASCB Cell Bio 2024
San Diego Convention
Center
San Diego, CA
December 14th-18th

Science and Art: Bridging
Two Creative Universes
(ASCB)
San Diego Convention
Center, CA
December 17th

Cytoskeleton Products

Actin Proteins
Activation Assays
Antibodies
ECM Proteins
ELISA Kits
G-LISA® Kits
Live Cell Imaging
Pull-down Assays
Motor Proteins
Small G-Proteins
Tubulin & FtsZ Proteins

Contact Us

P: 1 (303) 322.2254
F: 1 (303) 322.2257
E: cserve@cytoskeleton.com
W: cytoskeleton.com

For More News

W: cytoskeleton.com/blog

How the RhoA/ROCK Pathway Changes with Age and its Connection to the Diseases of Aging

Introduction

The Rho GTPases form a family of 20 closely related proteins with essential roles spanning diverse cellular processes. First known for their regulation of actin organization and dynamics, they are now also recognized as key mediators of gene expression and cell cycle progression¹.

Transforming protein RhoA, also known as Ras homolog family member A, was the first example to be discovered. Like other small GTPases, it serves as a “molecular switch” based on GTP/GDP binding status and is regulated by many other factors including guanine nucleotide exchange factors (GEFs), GTPase activating proteins (GAPs), and guanine nucleotide dissociation inhibitors (GDIs), through mechanisms that are reviewed in depth elsewhere².

RhoA signaling is propagated by an array of downstream effectors (Figure 1), the best characterized of which are the Rho-associated protein kinases, ROCK1 and ROCK2. In common with all Rho GTPases, RhoA contains a unique “insert domain” enabling direct interaction with these proteins³. When activated, ROCKs phosphorylate downstream targets including the PPP1R12A/MYPT1 subunit of myosin light chain phosphatase (MLCP), resulting in activation of MYL2/MLC2, the regulatory light chain of myosin II⁴. This effects actomyosin contractility, an underlying mechanism of force generation in cell motility and muscle contraction⁵. In addition, ROCKs can phosphorylate PPP1R14A/CPI-17, an inhibitor of MLCP⁶, and activate LIM domain kinases (LIMK1/2) toward cofilin phosphorylation to stabilize actin filaments⁷.

Ever since RhoA was proposed as an oncogene in the 1980s⁸ RhoA/ROCK signaling has been extensively studied in cancer and has been found to be dysregulated in many cancer types^{9,10}. However, this newsletter will focus on age-related changes that have also been identified across a range of studies, suggesting a role in other diseases of aging.

Age-Associated Variations in the RhoA/ROCK Pathway

These changes have mostly been studied in the cardiovascular system, as RhoA/ROCK signaling is one of two canonical pathways of muscle contraction (the other being myosin light chain kinase activation by calmodulin) and ROCKs are known to control vascular tone¹¹.

Arterial RhoA levels were first observed to increase with age¹², then a role was established for ROCK activation in declining contractile function¹³. In mice, age-dependent increases in arterial ROCK2 expression were functionally associated with increased myogenic tone (i.e., reduced capacity of blood vessel smooth muscle cells to contract/constrict in response to pressure changes)¹⁴. Thus, overactivation of RhoA and/or ROCKs causes prolonged contraction, potentially leading to vascular dysfunction that manifests as age-associated hypertension or arterial stiffening^{15,16}. Recent experiments with heterozygous knockout mice revealed a role for both ROCK isoforms in age-related aortic stiffening, with an apparently greater role for ROCK2¹⁶.

Increased expression of ROCK2 with age has also been found in the CNS¹⁷, where similarly, hippocampal RhoA protein levels and

GTPase activity are higher in old mice¹⁸. Another study reported both elevated RhoA expression and ROCK activity in the substantia nigra of older versus younger rats, and interestingly, these effects were attenuated by regular physical exercise¹⁹. The RhoA/ROCK pathway also affects neurogenesis, since progenitors derived from neural stem cells move through the ventricular-subventricular zone during lineage progression. Motility declines with age, impacting regenerative ability. One recent study used single-cell RNA-Seq to identify RhoA/ROCK signaling as a major contributing factor and found that pharmacological ROCK inhibition restored “young” progenitor cell dynamics²⁰.

With the above in mind, we will now briefly summarize current knowledge on RhoA/ROCK activation in some of the major diseases of aging: cardiovascular diseases, neurodegenerative diseases, and diabetes.

RhoA/ROCK Overactivation in Cardiovascular Diseases

As alluded to earlier, abnormal activation of RhoA/ROCK signaling has long been known in various cardiovascular conditions including hypertension, atherosclerosis, restenosis, and cardiac hypertrophy²¹. Higher pathway activity has also been observed in heart failure patients¹⁵. Several studies have linked age-related hypertension to RhoA/ROCK upregulation^{15,16,22}, and this has been further attributed to increased signaling via Gαq/11 proteins acting upstream of RhoA²³. A notable study in mice identified age-associated increases in arterial ROCK2 expression and basal myogenic tone. The latter could be markedly reduced by treatment with belumosudil, an FDA-approved ROCK2 inhibitor¹⁴.

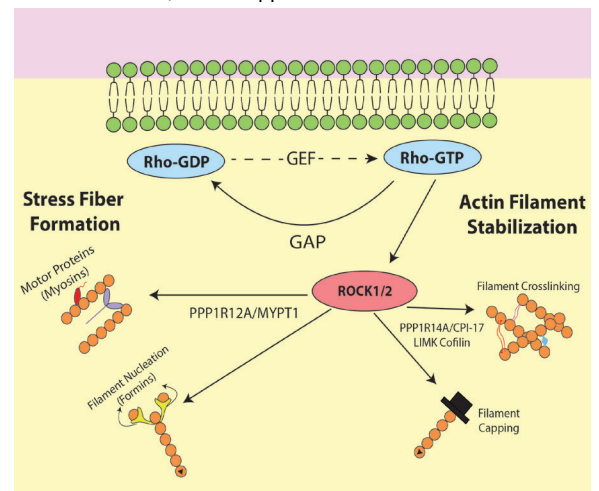


Figure Legend: Prenylated RhoA protein is closely associated with the plasma membrane, where GEFs and GAPs modulate the nucleotide status of RhoA. The GTP-form of RhoA activates ROCK1 and ROCK2 to modulate downstream effectors which result in stress fiber formation and actin filament growth and stabilization.



Helping advance science,
one protein at a time.

Continued from Page 1

Multiple Mechanisms in Neurodegeneration

While RhoA has normal biological roles in controlling axonal elongation and synaptic plasticity²⁴, dysregulation of the RhoA/ROCK pathway has been directly implicated in neurodegenerative disorders including Alzheimer's disease (AD), Parkinson's disease, and amyotrophic lateral sclerosis. Several pathogenic mechanisms have been proposed in the case of the former²⁵. First, heightened ROCK expression in AD brain can directly contribute to hyperphosphorylation of tau, potentiating the formation of neurofibrillary tangles that have also been shown to colocalize with RhoA²⁴. Second, ROCK activation and Aβ₁₋₄₀ secretion appear to amplify each other in a positive feedback loop. Additionally, increased actin contractility mediated by ROCK2 leads to neuronal dendritic spine loss, known as the major correlate of cognitive decline in AD²⁶.

Emerging Roles for RhoA/ROCK Signaling in Diabetes

Early studies²⁷ noted pathogenic increases in RhoA activity and phosphorylated CPI-17 (a direct ROCK substrate, as noted above) in aortas and arteries of db/db mice, a widely used animal model of type 2 diabetes (T2D). It is now established that RhoA mediates the translocation of SLC2A4/GLUT4, an essential transporter for glucose uptake into skeletal muscle²⁸, and that RhoA/ROCK1 regulate glucose transport in muscle cells and adipocytes through actin cytoskeleton remodeling²⁹. Thus, circulating insulin activates RhoA in muscle³⁰ while in pancreatic β cells, hyperglycemia upregulates RhoA/ROCK signaling to cause enhanced growth of stress fibers and reduced glucose-stimulated insulin secretion²⁹. Hyperactivation of RhoA therefore seems to contribute to diabetic pathology to the extent that RhoA/ROCK signaling was recently proposed as a novel therapeutic target in T2D, based on the available evidence²⁹.

Outstanding Questions and Future Prospects

Despite three decades of intensive study, several aspects of RhoA/ROCK signaling remain unclear. The precise functional roles of RhoA are still debated³¹ and some of the relevant mechanisms are not well understood due to inconclusive or contradictory results³². For example, while ROCK activation has been observed in heart failure patients¹⁵, cardiomyocyte-specific knockout of RhoA in mice led to accelerated heart aging and fibrosis³³. In a similar vein, studies in cancer suggest that RhoA can exert either pro- or anti-tumorigenic effects depending on the context¹⁰, highlighting how RhoA signaling networks may be tissue or cell type specific. Failure to consider different proteoforms could also be a confounding factor, given the many known posttranslational modification sites in RhoA that can modulate its activity and subcellular localization^{34,35}. Also, intriguing recent studies suggest RhoA is not predominantly membrane-bound as traditionally assumed and that it cycles much more rapidly between the membrane and cytoplasm than other Rho GTPases³¹. Nevertheless, the recent approval of belumosudil (a first-in-class selective ROCK2 inhibitor) for treatment of chronic graft-versus-host disease represents a notable success story, and the ongoing development of more potent and selective ROCK inhibitors offers new therapeutic potential for successfully targeting the RhoA/ROCK pathway in cancer⁹.

Rho, Rac, and Cdc42 Activation Assays

Product	Amount	Cat #
RhoA / Rac1 / Cdc42 Activation Assay Combo Biochem Kit	3 x 10 Assays	BK030
RhoA Pull-down Activation Assay Biochem Kit	80 Assays 20 Assays	BK036 BK036-S
RhoA G-LISA™ Activation Assay (Luminescence format)	96 Assays	BK121
RhoA G-LISA™ Activation Assay Kit (Colorimetric format)	96 Assays 24 Assays	BK124 BK124-S
Total RhoA ELISA	96 Assays	BK150
Rac1,2,3 G-LISA™ Activation Assay (Colorimetric format)	96 Assays	BK125
Rac1 G-LISA™ Activation Assay (Luminescence format)	96 Assays	BK126
Rac1 G-LISA™ Activation Assay Kit (Colorimetric Based)	96 Assays 24 Assays	BK128 BK128-S
Ras Pull-Down Activation Assay Biochem Kit	50 Assays	BK008
Ras G-LISA™ Activation Assay Kit (Colorimetric Based)	96 Assays	BK131

References

- Gray JL, von Delft F, Brennan PE. Targeting the small GTPase superfamily through their regulatory proteins. *Angew Chem Int Ed Engl.* 2020;59(16):6342–6366. <https://doi.org/10.1002/anie.201900585>.
- Cherfils J, Zeghouf M. Regulation of small GTPases by GEFs, GAPs, and GDIs. *Physiol Rev.* 2013;93(1):269–309. <https://doi.org/10.1152/physrev.00003.2012>.
- Zong H, Kaibuchi K, Quilliam LA. The insert region of RhoA is essential for Rho kinase activation and cellular transformation. *Mol Cell Biol.* 2001;21(16):5287–5298. <https://doi.org/10.1128/mcb.21.16.5287-5298.2001>.
- Pandya P, Orgaz JL, Sanz-Moreno V. Actomyosin contractility and collective migration: May the force be with you. *Curr Opin Cell Biol.* 2017;48:87–96. <https://doi.org/10.1016/j.ccb.2017.06.006>.
- Murrell M, Oakes PW, Lenz M, Gardel ML. Forcing cells into shape: The mechanics of actomyosin contractility. *Nat Rev Mol Cell Biol.* 2015;16(8):486–498. <https://doi.org/10.1038/nrm4012>.
- Loirand G. Rho kinases in health and disease: From basic science to translational research. *Pharmacol Rev.* 2015;67(4):1074–1095. <https://doi.org/10.1124/pr.115.010595>.
- Lu W, Wen J, Chen Z. Distinct roles of ROCK1 and ROCK2 on the cerebral ischemia injury and subsequently neurodegenerative changes. *Pharmacology.* 2020;105(1–2):3–8. <https://doi.org/10.1159/000502914>.
- Gilbert-Ross M, Marcus AJ, Zhou W. RhoA, a novel tumor suppressor or oncogene as a therapeutic target? *Genes Dis.* 2015;2(1):2–3. <https://doi.org/10.1016/j.gendis.2014.10.001>.
- Barcelo J, Samain R, Sanz-Moreno V. Preclinical to clinical utility of ROCK inhibitors in cancer. *Trends Cancer.* 2023;9(3):250–263. <https://doi.org/10.1016/j.trecan.2022.12.001>.
- Santos JC, Proffitts-Pelejón N, Sánchez-Vinces S, Roué G. RHOA therapeutic targeting in hematological cancers. *Cells.* 2023;12(3):433. <https://doi.org/10.3390/cells12030433>.
- Barbosa GS, Costa RM, Awata WMC, et al. Suppressed vascular Rho-kinase activation is a protective cardiovascular mechanism in obese female mice. *Biosci Rep.* 2023;43(7):BSR20230672. <https://doi.org/10.1042/bsr20230672>.
- Miao L, Calvert JW, Tang J, Parent AD, Zhang JH. Age-related RhoA expression in blood vessels of rats. *Mech Ageing Dev.* 2001;122(15):1757–1770. [https://doi.org/10.1016/S0047-6374\(01\)00297-4](https://doi.org/10.1016/S0047-6374(01)00297-4).
- Seawright JW, Sreenivasappa H, Gibbs HC, et al. Vascular smooth muscle contractile function declines with age in skeletal muscle feed arteries. *Front Physiol.* 2018;9:856. <https://doi.org/10.3389/fphys.2018.00856>.
- Björling K, Joseph PD, Egebjerg K, Salomonsson M, Hansen LJ, Ludvigsen TP, Jensen LJ. Role of age, Rho-kinase 2 expression, and G protein-mediated signaling in the myogenic response in mouse small mesenteric arteries. *Physiol Rep.* 2018;6(17):e13863. <https://doi.org/10.14814/phyz.13863>.
- Palomo I, Wehinger S, Andrés V, García-García FJ, Fuentes E. RhoA/rho kinase pathway activation in age-associated endothelial cell dysfunction and thrombosis. *J Cell Mol Med.* 2024;28(8):e18153. <https://doi.org/10.1111/jcmm.18153>.
- Li Y, Tai H-C, Sladojevich N, Kim H-H, Liao JK. Vascular stiffening mediated by Rho-associated coiled-coil containing kinase isoforms. *J Am Heart Assoc.* 2021;10(20):e022568. <https://doi.org/10.1161/jaha.121.022568>.
- Tonges L, Koch J-C, Bähr M, Lingor P. ROCKing regeneration: Rho kinase inhibition as molecular target for neurorestoration. *Front Mol Neurosci.* 2011;4:39. <https://doi.org/10.3389/fnmol.2011.00039>.
- Wong L-K, Chong YS, Lin W, Kisiswa L, Sim E, Ibañez CF, Sajikumar S. Age-related changes in hippocampal-dependent synaptic plasticity and memory mediated by p75 neurotrophin receptor. *Aging Cell.* 2021;20(2):e13305. <https://doi.org/10.1111/acel.13305>.
- Muñoz A, Corría CL, Villar-Cheda B, Costa-Besada MA, Labandeira-García JL. Aging-related increase in Rho kinase activity in the nigral region is counteracted by physical exercise. *J Gerontol A Biol Sci Med Sci.* 2016;71(10):1254–1257. <https://doi.org/10.1093/gerona/glv179>.
- Zhao X, Fisher ES, Wang Y, Zuloga K, Manley L, Temple S. 4D imaging analysis of the aging mouse neural stem cell niche reveals a dramatic loss of progenitor cell dynamics regulated by the RHO-ROCK pathway. *Stem Cell Reports.* 2022;17(2):245–258. <https://doi.org/10.1016/j.stemcr.2021.12.007>.
- Loirand G, Guérin P, Pacaud P. Rho kinases in cardiovascular physiology and pathophysiology. *Circ Res.* 2006;98(3):322–334. <https://doi.org/10.1161/01.res.0000201960.04223.3c>.
- Rabaglino MB, Wakabayashi M, Pearson JT, Jensen LJ. Effect of age on the vascular proteome in middle cerebral arteries and mesenteric resistance arteries in mice. *Mech Ageing Dev.* 2021;200:111594. <https://doi.org/10.1016/j.mad.2021.111594>.
- Nunes KP, Webb RC. New insights into RhoA/Rho-kinase signaling: A key regulator of vascular contraction. *Small GTPases.* 2021;12(5–6):458–469. <https://doi.org/10.1080/21541248.2020.1822721>.
- Akhtar SN, Bunner WP, Brennan E, Lu Q, Sztamari EM. Crosstalk between the Rho and Rab family of small GTPases in neurodegenerative disorders. *Front Cell Neurosci.* 2023;17:1084769. <https://doi.org/10.3389/fncel.2023.1084769>.
- Sastre AA, Montoro ML, Gálvez-Martín P, Lacerda HM, Lucía AM, Llaveró F, Zugaza JL. Small GTPases of the Ras and Rho families switch on/off signaling pathways in neurodegenerative diseases. *Int J Mol Sci.* 2020;21(17):6312. <https://doi.org/10.3390/ijms21176312>.
- Walker CK, Herszkowitz JH. Dendritic spines: Mediators of cognitive resilience in aging and Alzheimer's disease. *Neuroscientist.* 2021;27(5):487–505. <https://doi.org/10.1177/1073858420945964>.
- Xie Z, Su W, Guo Z, Pang H, Post SR, Gong MC. Up-regulation of CPI-17 phosphorylation in diabetic vasculature and high glucose cultured vascular smooth muscle cells. *Cardiovasc Res.* 2006;69(2):491–501. <https://doi.org/10.1016/j.cardiores.2005.11.002>.
- Møller LLL, Klip A, Saylor L. Rho GTPases—emerging regulators of glucose homeostasis and metabolic health. *Cells.* 2019;8(5):434. <https://doi.org/10.3390/cells8050434>.
- Gendaszewska-Darmach E, Garstka M, Błażewska KM. Targeting small GTPases and their prenylation in diabetes mellitus. *J Med Chem.* 2021;64(14):9677–9710. <https://doi.org/10.1021/acs.jmedchem.1c00410>.
- Machin PA, Tsounou E, Hornigold DC, Welch HCE. Rho family GTPases and Rho GEFs in glucose homeostasis. *Cells.* 2021;10(4):915. <https://doi.org/10.3390/cells10040915>.
- de Seze J, Gatlin J, Coppey M. RhoA regulation in space and time. *FEBS Lett.* 2023;597(6):836–849. <https://doi.org/10.1002/1873-3468.14578>.
- Kilian LS, Voran J, Frank D, Rangrez AY. RhoA: A dubious molecule in cardiac pathophysiology. *J Biomed Sci.* 2021;28(1):33. <https://doi.org/10.1186/s12929-021-00730-w>.
- Soh JEC, Shimizu A, Molla MR, et al. RhoA rescues cardiac senescence by regulating Parkin-mediated mitophagy. *J Biol Chem.* 2023;298(3):102993. <https://doi.org/10.1016/j.jbc.2023.102993>.
- Navarro-Lérida I, Sánchez-Álvarez M, Del Pozo MÁ. Post-translational modification and subcellular compartmentalization: Emerging concepts on the regulation and pathophysiological relevance of Rho GTPases. *Cells.* 2021;10(8):1990. <https://doi.org/10.3390/cells10081990>.
- Schmidt SI, Bjaalberg M, Freude K, Meyer M. RhoA signaling in neurodegenerative diseases. *Cells.* 2022;11(9):1520. <https://doi.org/10.3390/cells11091520>.

RhoA Small G-Proteins, Activators and Inhibitors

Labeled Actin	Amount	Cat. #
RhoA Protein: His Tagged Human Wild Type	1 x 100 µg 3 x 100 µg 1 x 1 mg	RH01-A RH01-C RH01-XL
RhoA Protein: His Tagged: Human Constitutively Active	1 x 10 µg	R6301-A
Rho Activator I	5 x 10 units 20 x 10 units	CN01-A CN01-B
Rho Activator II	3 x 20 µg 9 x 20 µg	CN03-A CN03-B
Rho Inhibitor I	1 x 20 µg 5 x 20 µg 20 x 20 µg	CT04-A CT04-B CT04-C