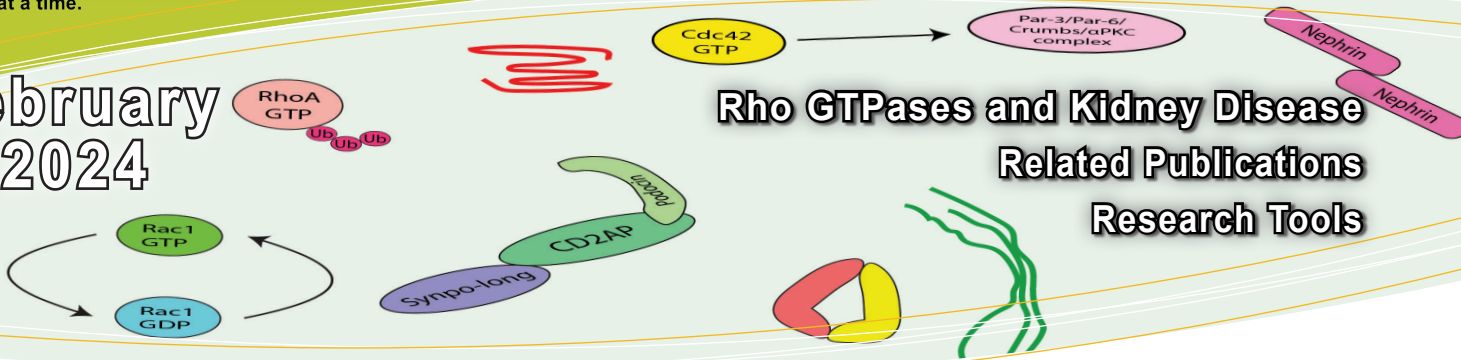




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February 2024

Rho GTPases and Kidney Disease Related Publications Research Tools



Rho GTPases and Kidney Disease

Introduction

The Rho family of small GTPases are molecular switches with a pivotal role in numerous cellular functions. They regulate the actin cytoskeleton, cellular morphology, motility, adhesion, and proliferation^{1,2}. RhoA, Rac1, and Cdc42 are members of the Rho family that play crucial roles in kidney function, particularly in the maintenance of various kidney cell types, such as tubular epithelial cells, mesangial cells, and podocytes³ (Figure 1). Podocytes are a fundamental component of the glomerular filtration barrier, a structure that prevents the filtration of large proteins and macromolecules into the urine. This glomerular barrier relies on the structural integrity of the actin-based cytoskeleton of the podocytes⁴. As a result, dysregulation of GTPases and their downstream effectors has been implicated in both hereditary and idiopathic kidney diseases. The present newsletter will dive into Rho family-dependent nephropathies and their underlying molecular mechanisms.

Rho GTPases and diabetic nephropathies

Diabetic nephropathy, a progressive kidney disease that arises from glomerular injury, is linked to RhoA activation in mesangial cells under the influence of high glucose levels, ultimately resulting in excessive extracellular matrix production (glomerulosclerosis)⁵. The pathogenesis of diabetic nephropathy is a complex interplay of factors, including hyperglycemia, advanced glycation end products, oxidative stress, and the activation of various cellular pathways. These elements stimulate Rho-GTPase activity, initiating a cascade that culminates in renal damage^{6,7}. Additionally, Cdc42 is involved in the regulation of insulin synthesis, insulin granule mobilization, and exocytosis⁸, while Rac1 influences oxidative damage and apoptotic activity in mesangial cells^{9,10}.

Rho GTPases and congenital nephropathies

Rho GTPases play a crucial role in the landscape of congenital nephropathies, notably influencing Focal Segmental Glomerulo-Sclerosis (FSGS), Autosomal Dominant Polycystic Kidney Disease (ADPKD), and Alport syndrome, among others. The nature of these renal conditions and their associations to Rho GTPases will be discussed below.

FSGS describes a histological pattern of disease observed in kidney biopsy, with a phenotype defined by scar tissue in the glomerulus. As such, it can serve as a diagnosis indicator representing various injuries sustained by the kidneys¹¹. Initially documented in 1925 among adults exhibiting nephritic proteinuria¹², FSGS can manifest

as idiopathic, genetic, autoimmune, or as a response (maladaptive or otherwise) to chronic kidney disease associated with reduced nephron mass, as well as secondary to drugs or infections¹¹. In both inherited and sporadic forms of FSGS, mutations in genes encoding regulators of Rho GTPase activities have been identified. These mutations involve the GTPase-activating protein (GAP) Rho-GAP 24 (ARHGAP24), which acts as a GAP for Rac1 and controls the RhoA-Rac1 signaling balance^{11,13,14}. Additionally, mutations in Rho GDP Dissociation Inhibitor alpha (ARHGDIa) and intersectin-1 and -2 (ITSN-1 and -2) have been linked to congenital nephrotic syndrome¹⁵⁻¹⁷. Notably, mutations in ARHGAP24 lead to excessive Rac1 signaling in podocytes, while mutations in ARHGDIa result in the loss of function of RhoGDIa, a negative Rho regulator, causing hyperactivation of Rho-GTPases and impaired cell motility.

ADPKD stands as the most prevalent hereditary kidney ailment and ranks among the leading contributors to end-stage kidney disease. The clinical manifestations of ADPKD involve enlarged kidneys housing proliferating cysts, hypertension, and various extrarenal complications^{18,19}. The majority of cases (>90%) are attributed to mutations in two genes, namely PKD1 and PKD2²⁰. The cellular phenotype associated with ADPKD is complex and includes changes in proliferation, apoptosis, cell-cell and cell-matrix adhesion, among other manifestations²¹. In this regard, a mutation in PKD1 was recently found to induce substantial alterations in the dynamics of the actin cytoskeleton. Particularly, the study revealed that

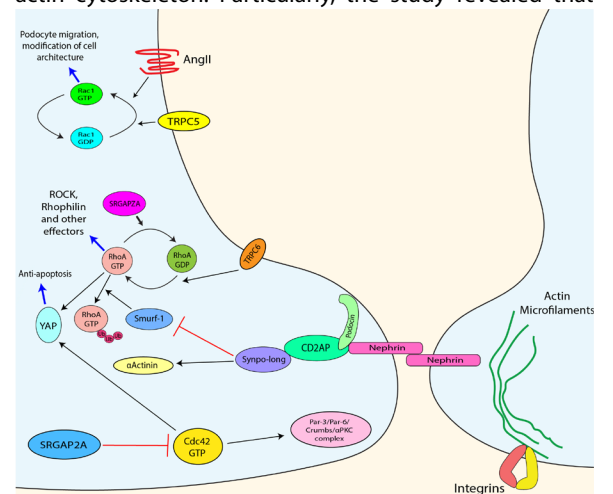


Figure 1. Schematic representation of maintenance of podocyte architecture by Rho GTPases. Adapted from Steichen et al. Small GTPases 2022.

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the dysregulation of compartmentalized centrosomal RhoA signaling, facilitated by a particular Rho GTPase-activating protein (ARHGAP35), resulted in heightened activation of Rho kinase (ROCK) in cells with a PKD1 mutation. In turn, there was an increase in F-actin polymerization and contractility²². Furthermore, a crucial signaling pathway involving the Rho guanine nucleotide exchange factor (GEF) LARG, RhoA, its effector ROCK, and myosin light chain (MLC) was identified as a critical mechanism for PKD1 effects in ADPKD²³.

Alport syndrome is a rare hereditary glomerulonephritis associated with progressive loss of kidney function, hearing loss and eye abnormalities²⁴. A mechanistic connection between Rho GTPases and the development of Alport syndrome has been previously established. The stretch-induced activation of Rac1 and Cdc42 serves as a trigger for mesangial filopodial invasion within the glomerular capillary loops²⁵. Recently, a further indirect relationship between this syndrome and Rac1 and Cdc42 was found through the involvement of the Rac1/Cdc42 guanine nucleotide exchange factor β 1Pix²⁶. According to this study, β 1Pix plays a role in regulating LDL receptor stability and function in renal cells affected by this condition, with implications in its pathogenesis.

Summary and future perspectives

Efficient kidney barrier function necessitates the dynamic regulation of the podocyte cytoskeleton. Rho GTPases, as master regulators of actin cytoskeletal dynamics, are thus critically important for maintaining the sustained function of the kidney barrier. These proteins play crucial roles in congenital nephropathies like FSGS, ADPKD, and Alport syndrome. Additionally, Rho GTPases are known for their role in HIV-associated nephropathy, an ailment characterized by collapsing focal segmental glomerulosclerosis (cFSGS)²⁷. In this context, Nef, the HIV-1 accessory and determinant protein, influences podocyte morphology by inhibiting RhoA activity and activating Rac1. Moreover, HIV-1 transactivator Tat, combined with FGF-2, induces podocyte dedifferentiation and proliferation through the stimulation of the RhoA/phospho-Myosin Light Chain 2 pathways^{28,29}. Further research has identified host proteins interacting with Nef, among which is TNK2, also known as ACK, closely involved with Cdc42 activity³⁰. Overall, Rho GTPases are master regulators of the actin cytoskeleton and their dysregulation is implicated in the pathogenesis of glomerular diseases. However, given the diverse biological functions associated with Rho GTPases, it is imperative to thoroughly investigate potential therapeutic targets while also identifying signaling pathways that minimize 'off-target' effects.

G-LISA Activation Assay Kits

Product	Assays	Cat. #
Cdc42 G-LISA™ Activation Assay Kit (Colorimetric format)	96	BK127
Rac1 G-LISA™ Activation Assay Kit (Luminescence format)	96	BK126
Rac1 G-LISA™ Activation Assay Kit (Colorimetric format)	96	BK128
Ra1A G-LISA™ Activation Assay Kit (Colorimetric format)	96	BK129
RhoA G-LISA™ Activation Assay (Luminescence format)	96	BK121
RhoA G-LISA™ Activation Assay Kit (Colorimetric format)	96	BK124
RhoA / Rac1/ Cdc42 G-LISA™ Activation Assay Bundle 3 Kits (24 assays per kit)	96	BK135
Rac1,2,3 G-LISA™ Activation Assay (Colorimetric format)	96	BK125
Rac1 G-LISA™ Activation Assay (Luminescence format)	96	BK126
Rac1 G-LISA™ Activation Assay Kit (Colorimetric Based)	96	BK128
Ras G-LISA™ Activation Assay Kit (Colorimetric Based)	96	BK131
Total RhoA ELISA	96	BK150

References

- Foxman, E., Ibrahim, S. & Takano, T. Rho GTPase regulatory proteins contribute to podocyte morphology and function. *McGill Sci. Undergrad. Res. J.* 18, A11–A17 (2023).
- Lin, Y. & Zheng, Y. Rho family GTPases and their regulators. in *NADPH Oxidases Revisited: From Function to Structure 287* (Springer Nature, 2023).
- Liang, H. et al. Pharmacological inhibition of Rac1 exerts a protective role in ischemia/reperfusion-induced renal fibrosis. *Biochem. Biophys. Res. Commun.* 503, 2517–2523 (2018).
- Blaine, J. & Dylewski, J. Regulation of the Actin Cytoskeleton in Podocytes. *Cells* 9, (2020).
- Kravets, I. & Mallipattu, S. K. The Role of Podocytes and Podocyte-Associated Biomarkers in Diagnosis and Treatment of Diabetic Kidney Disease. *J. Endocr. Soc.* 4, (2020).
- Lin, J. S. & Susztak, K. Podocytes: the Weakest Link in Diabetic Kidney Disease? *Curr. Diab. Rep.* 16, 45 (2016).
- Kawanami, D., Matoba, K. & Utsunomiya, K. Signaling pathways in diabetic nephropathy. *Histol. Histopathol.* 31, 1059–67 (2016).
- Huang, Q.-Y. et al. Cdc42: A Novel Regulator of Insulin Secretion and Diabetes-Associated Diseases. *Int. J. Mol. Sci.* 20, (2019).
- Lin, C.-L. et al. Superoxide destabilization of beta-catenin augments apoptosis of high-glucose-stressed mesangial cells. *Endocrinology* 149, 2934–42 (2008).
- Tung, C., Hsu, Y., Shih, Y., Chang, P. & Lin, C. Glomerular mesangial cell and podocyte injuries in diabetic nephropathy. *Nephrology* 23, 32–37 (2018).
- Sambharia, M., Rastogi, P. & Thomas, C. P. Monogenic focal segmental glomerulosclerosis: A conceptual framework for identification and management of a heterogeneous disease. *Am. J. Med. Genet. Part C Semin. Med. Genet.* 190, 377–398 (2022).
- Cameron, J. S. Focal segmental glomerulosclerosis in adults. *Nephrol. Dial. Transplant.* 18, vi45–vi51 (2003).
- Akilesh, S. et al. Arhgap24 inactivates Rac1 in mouse podocytes, and a mutant form is associated with familial focal segmental glomerulosclerosis. *J. Clin. Invest.* 121, 4127–4137 (2011).
- Daehn, I. S. & Duffield, J. S. The glomerular filtration barrier: a structural target for novel kidney therapies. *Nat. Rev. Drug Discov.* 20, 770–788 (2021).
- Najafi, M. et al. High detection rate for disease-causing variants in a cohort of 30 Iranian pediatric steroid resistant nephrotic syndrome cases. *Front. Pediatr.* 10, (2022).
- Gee, H. Y. et al. ARHGDI4 mutations cause nephrotic syndrome via defective RHO GTPase signaling. *J. Clin. Invest.* 123, 3243–53 (2013).
- Ashraf, S. et al. Mutations in six nephrosis genes delineate a pathogenic pathway amenable to treatment. *Nat. Commun.* 9, 1960 (2018).
- Cornec-Le Gall, E., Alam, A. & Perrone, R. D. Autosomal dominant polycystic kidney disease. *Lancet* 393, 919–935 (2019).
- Ong, A. C. M., Devuyst, O., Knebelmann, B., Walz, G. & ERA-EDTA Working Group for Inherited Kidney Diseases. Autosomal dominant polycystic kidney disease: the changing face of clinical management. *Lancet (London, England)* 385, 1993–2002 (2015).
- Ong, A. C. M. Making sense of polycystic kidney disease. *Lancet (London, England)* 389, 1780–1782 (2017).
- Ong, A. C. M. & Harris, P. C. A polycystin-centric view of cyst formation and disease: the polycystins revisited. *Kidney Int.* 88, 699–710 (2015).
- Streets, A. J., Prosseda, P. P. & Ong, A. C. M. Polycystin-1 regulates ARHGAP35-dependent centrosomal RhoA activation and ROCK signaling. *JCI Insight* 5, (2020).
- Cai, J. et al. A RhoA-YAP-c-Myc signaling axis promotes the development of polycystic kidney disease. *Genes Dev.* 32, 781–793 (2018).
- Mahrous, N. N. et al. A Current Landscape on Alport Syndrome Cases: Characterization, Therapy and Management Perspectives. *Biomedicines* 11, 2762 (2023).
- Zalocchi, M., Johnson, B. M., Meehan, D. T., Delimont, D. & Cosgrove, D. α 1 β 1 integrin/Rac1-dependent mesangial invasion of glomerular capillaries in Alport syndrome. *Am. J. Pathol.* 183, 1269–1280 (2013).
- Chahdi, A. et al. β Pix sequesters IDOL and prevents LDL receptor degradation through a β 2AR-regulated signaling pathway in Alport Syndrome. *Prepr. from bioRxiv* (2020). doi:10.1101/2020.11.06.372292
- Chen, A., Yin, L., Lee, K. & He, J. C. Similarities and Differences between COVID-19-Associated Nephropathy and HIV-Associated Nephropathy. *Kidney Dis.* 8, 1–12 (2022).
- Matsuda, J., Asano-Matsuda, K., Kitzler, T. M. & Takano, T. Rho GTPase regulatory proteins in podocytes. *Kidney Int.* 99, 336–345 (2021).
- Lu, T. et al. HIV-1 Nef Disrupts the Podocyte Actin Cytoskeleton by Interacting with Diaphanous Interacting Protein. *J. Biol. Chem.* 283, 8173–8182 (2008).
- Kim, J.-A., Shin, Y.-H. & Yoon, C.-H. The HIV-1 Virulence Factor Nef as a New Therapeutic Target Against HIV/AIDS. *J. Bacteriol. Virol.* 50, 187–194 (2020).

RhoA Small G-Proteins, Activators and Inhibitors

Labeled Actin	Amount	Cat. #
RhoA Protein: His Tagged Human Wild Type	1 x 100 μ g 3 x 100 μ g 1 x 1 mg	RH01-A RH01-C RH01-XL
RhoA Protein: His Tagged: Human Constitutively Active	1 x 10 μ g	R6301-A
Rho Activator I	5 x 10 units 20 x 10 units	CN01-A CN01-B
Rho Activator II	3 x 20 μ g 9 x 20 μ g	CN03-A CN03-B
Rho Inhibitor I	1 x 20 μ g 5 x 20 μ g 20 x 20 μ g	CT04-A CT04-B CT04-C